

15-OXO-ZOAPATLIN, A DITERPENE LACTONE FROM *VIGUIERA MACULATA*

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Abstract—The isolation of a new diterpene lactone and two known diterpenoid acids from the aerial parts of *Viguiera maculata* is reported

INTRODUCTION

Chemical examinations of the large genus *Viguiera* (Compositae, Heliantheae) have yielded diterpenes [1–3], sesquiterpene lactones [4–10] and flavonol compounds [11]. We now report the isolation and structure determination of 15-oxo-zoapatlin (1), a new diterpene lactone found as a constituent of *Viguiera maculata* Blake. This plant also contains the known diterpenoid acids, *ent*-kaur-16-en-19-oic (2) and 15 α -hydroxy-*ent*-kaur-16-en-19-oic (3) [13] by their physical constants and direct comparison with authentic samples.

RESULTS AND DISCUSSION

Chromatographic separation of a chloroform extract of the aerial parts of *Viguiera maculata* afforded three diterpenoid compounds. The two most abundant were identified as the known compounds *ent*-kaur-16-en-19-oic acid (2) [12] and 15 α -hydroxy-*ent*-kaur-16-en-19-oic acid (3) [13] by their physical constants and direct comparison with authentic samples.

The third diterpene, 15-oxo-zoapatlin (1) C₂₀H₂₆O₃, contained an α,β -unsaturated ketone (UV λ_{\max} 233 nm, ϵ 6083) conjugated with an exocyclic methylene group (¹H NMR δ 5.95 t, J = 1 Hz, 5.20 t, J = 1 Hz). The ketone was located on a cyclopentane ring (IR 1718, 1639 cm⁻¹). These data clearly indicate the nature of the D ring of a tetracyclic diterpene. Two methyl singlets at δ 1.27 and 1.10 in the ¹H NMR spectrum suggested that this compound belonged to the kaurene or modified kaurene series. Since a γ -lactone (IR 1755 cm⁻¹, ¹³C NMR δ 180.27 s) was closed to a quaternary carbon (¹³C NMR δ 87.64 s), this new substance was a modified *ent*-kauranoid with the C-10 methyl group shifted to C-9, similar to eupatalbin and eupatoralbin [14]. It was therefore identified as the 15-oxo-derivative of zoapatlin (4) [15].

The ¹³C NMR spectrum of 15-oxo-zoapatlin is in complete agreement with the proposed structure and the assignments were established by comparison with the

spectra of various kauranoids [16] and closely related modified kauranoids [14, 17].

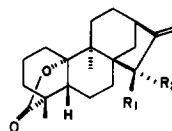
The fact that the new compound was indeed the 15-oxo-derivative of zoapatlin was confirmed by chemical correlation. Zoapatlin (4), still available from earlier work, was transformed to 1 by treatment of 4 with SeO₂ yielding the 15 α -hydroxy derivative 5, which was treated with CrO₃-pyridine to afford 15-oxo-zoapatlin, identical in all respects with the natural product.

Zoapatlin (4), first isolated from *Montanoa tomentosa* [15], was shown to be identical by direct comparison with the diterpene lactone tetrachyrin, isolated from *Tetrachyron orizabensis* var *websteri* and *Helianthus debilis* subsp *debilis* [17]. Therefore the name tetrachyrin should be abandoned.

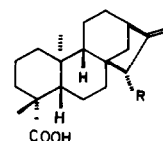
The results indicate a great similarity in chemical composition between *Viguiera* [1–11] and *Helianthus* [18–25] species since both genera of the subtribe Helianthinae elaborate closely related sesquiterpene lactones and diterpenoids.

EXPERIMENTAL

CHCl₃ extraction of 960 g of the above ground parts of *Viguiera maculata* Blake (voucher on deposit in the National Herbarium of Mexico, Instituto de Biología de la UNAM, Reg No 282569), collected 125 km SSE Izúcar de Matamoros, Puebla, afforded 34.7 g of crude gum. This was chromatographed over 1.2 kg of



- 1 $R_1, R_2 = O$
4 $R_1 = R_2 = H$
5 $R_1 = H, R_2 = OH$



- 2 $R = H$
3 $R = OH$

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silica gel using a hexane-EtOAc gradient elution system. All the fractions were monitored by TLC. Some fractions eluted with hexane-EtOAc (7/3), which showed the same spot on TLC, were combined to yield 1.9 g residue. This was rechromatographed over silica gel (60 g) and elution with hexane-EtOAc (9/1) afforded crude 15-oxo-zoapatlin (1), recrystallization from EtOAc-iso-Pr₂O yielded 347.3 mg of 1, mp 164–165°, $[\alpha]_D^{25}$ –74° (CHCl₃, c 0.123), IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹ 1755, 1715, 1690, 880; ¹H NMR (CDCl₃, 80 MHz) δ 5.94 (dd, *J* = 1 Hz, H-17a), 5.20 (dd, *J* = 1 Hz, H-17b), 2.87 (m, 1H, H-13), 1.27 (s, 3H, C-18), 1.10 (s, 3H, C-20), ¹³C NMR (CDCl₃, 20 MHz) δ 210.39 (s, C-15), 180.27 (s, C-19), 149.20 (s, C-16), 114.92 (t, C-17), 87.64 (s, C-10), 52.09 (s, C-8), 52.00 (d, C-5), 47.51 (s, C-9), 43.64 (s, C-4), 39.15 (t, C-1), 37.62 (d, C-13), 35.50 (t, C-14), 31.08 (t, C-7), 30.19 (t, C-3), 29.64 (t, C-12), 25.71 (t, C-11), 20.12 (t, C-6), 18.54 (q, C-20), 18.24 (t, C-2), 17.06 (q, C-18), MS (direct inlet) 75 eV, *m/z* (rel int) 314 [M]⁺ (86), 271 (100), 270 (56), 255 (39), 237 (18), 212 (34), 199 (22) (Found: C, 76.28, H, 8.36, O, 15.28%. C₂₀H₂₆O₃ requires C, 76.40, H, 8.34, O, 15.27%).

Subsequent fractions of the initial CC were combined affording 6.17 g residue, which was rechromatographed on silica gel (180 g) using hexane-EtOAc (9/1) as constant eluent. From this column, were isolated 1.6287 g of *ent*-kaur-16-en-19-oic acid (2), mp 178–180°, IR, ¹H NMR, ¹³C NMR and MS identical with authentic material [1, 17]. From the fractions eluted with hexane-EtOAc (3/2) of the initial CC, were isolated 17.4 mg of 15 α -hydroxy-*ent*-kaur-16-en-19-oic acid (3), mp 230–231° (lit 229–231° [11], 230–232° [20]), identical by direct comparison with an authentic sample.

Oxidation of zoapatlin. Compound 4 (75 mg) was treated with SeO₂ (15 mg) in dioxan (5 ml) and H₂O (1.5 ml) at room temp for 5 hr. Usual work-up yielded a residue which was chromatographed on silica gel (1 g) using hexane-EtOAc (4/1) as eluent, 44 mg of 15 α -hydroxyzoapatlin (5) were obtained. Mp 169–171°, IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm⁻¹ 3500, 1756, 1680, ¹H NMR (CDCl₃, 80 MHz) δ 5.21 (br s, *W*_{1/2} = 3 Hz, H-17a), 5.06 (br s, *W*_{1/2} = 3 Hz, H-17b), 4.08 (br s, *W*_{1/2} = 3 Hz, H-15), 2.55 (m, H-13), 1.19 (s, 3H, C-18), 1.08 (s, 3H, C-20), MS (direct inlet) 75 eV *m/z* (rel int) 316 [M]⁺ (91), 274 (53), 220 (69), 147 (39), 105 (62), 91 (100), 79 (68), 55 (48). Compound 5 (34 mg) was treated with CrO₃ (50 mg) in pyridine (1 ml) at 0° for 12 hr followed by the usual work-up, to yield, after purification through a small silica gel column (0.5 g), 25 mg of 1, IR, ¹H NMR, ¹³C NMR and MS identical with the natural product isolated from *Viguiera maculata*.

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REFERENCES

- Delgado, G., Romo de Vivar, A., Ortega, A., Cárdenas, J. and Schlemper, E. O. (1983) *Phytochemistry* **22**, 1227.
- Delgado, G., Romo de Vivar, A., Cárdenas, J., Pereda-Miranda, R. and Huerta, E. (1984) *Phytochemistry* **23**, 2285.
- Bohlmann, F., Zdero, C. and Mahanta, P. (1977) *Phytochemistry* **16**, 1073.
- Romo de Vivar, A., Guerrero, C., Díaz, E., Bratoeff, E. and Jiménez, L. (1976) *Phytochemistry* **15**, 525.
- Romo de Vivar, A., Delgado, G., Guerrero, C., Reséndiz, J. and Ortega, A. (1978) *Rev. Latinoam. Quím.* **9**, 171.
- Guerrero, C., Santana, M. and Romo, J. (1976) *Rev. Latinoam. Quím.* **7**, 41.
- Ortega, A., Lara, R., Martínez, R. and Díaz, E. (1980) *Phytochemistry* **19**, 1545.
- Romo de Vivar, A., Bratoeff, E., Ontiveros, E., Lankin, D. C. and Bhacca, N. S. (1980) *Phytochemistry* **19**, 1795.
- Delgado, G., Romo de Vivar, A. and Herz, W. (1982) *Phytochemistry* **21**, 1305.
- Bohlmann, F., Jakupovic, J., Ahmed, M., Grenz, M., Suding, H., Robinson, H. and King, R. M. (1981) *Phytochemistry* **20**, 113.
- Delgado, G., Alvarez, L. and Romo de Vivar, A. (1984) *Phytochemistry* **23**, 675.
- Henrick, C. A. and Jefferies, P. R. (1964) *Aust. J. Chem.* **17**, 915.
- Piozzi, F., Spiro, V., Passannanti, S. and Mondelli, R. (1968) *Gazz. Chim. Ital.* **98**, 907.
- Herz, W., Gorindan, S. and Blount, J. F. (1979) *J. Org. Chem.* **44**, 2999.
- Caballero, Y. and Walls, F. (1970) *Bol. Inst. Quím., Univ. Nac. Auton. Mex.* **22**, 79.
- Wehrli, F. W. and Nishida, T. (1979) *Fortschr. Chem. Org. Naturst.* **36**, 1.
- Ohno, N., Nabry, T. J., Zabel, V. and Watson, W. H. (1979) *Phytochemistry* **18**, 1687.
- Bjeldanes, L. F. and Geissman, T. A. (1970) *Phytochemistry* **11**, 327.
- Herz, W. and Kumar, N. (1981) *Phytochemistry* **20**, 99.
- Ohno, N. and Mabry, T. J. (1980) *Phytochemistry* **19**, 609.
- Bohlmann, F., Jakupovic, J., King, R. M. and Robinson, H. (1980) *Phytochemistry* **19**, 863.
- Herz, W. and Kumar, N. (1981) *Phytochemistry* **20**, 93.
- Ohno, N. and Mabry, T. J. (1979) *Phytochemistry* **18**, 1003.
- Herz, W., Govindan, S. V. and Watanabe, K. (1982) *Phytochemistry* **21**, 946.
- Herz, W., Kulanthiavel, P. and Watanabe, K. (1983) *Phytochemistry* **22**, 2021.